

## Genome Biology of Cancer

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## The birth of cancer genomics



Theodor Boveri

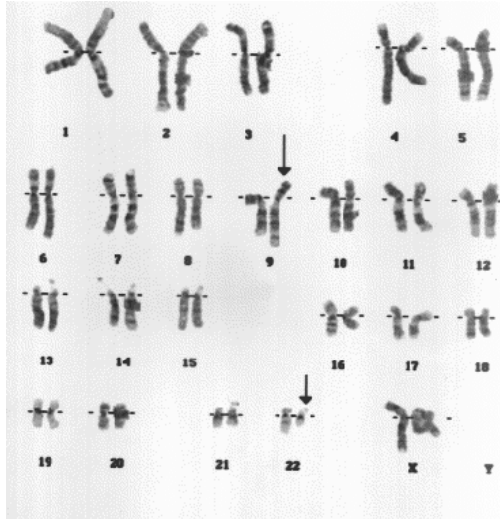
### Somatic mutation theory of cancer

Based on observations of abnormal growth of sea-urchin eggs that carry the “wrong” chromosomal complement, Boveri proposed that tumour growth is based on a similar, but particular, incorrect combination of chromosomes.



Boveri TH (1914). Zur Frage der Entstehung maligner Tumoren. Verlag von Gustav Fisher, Jena.

## Karyotype with the Philadelphia chromosome



Translocation between chromosomes 9 & 22 in a chronic myelogenous leukaemia (CML);  $t(9;22)(q34;q11)$ .

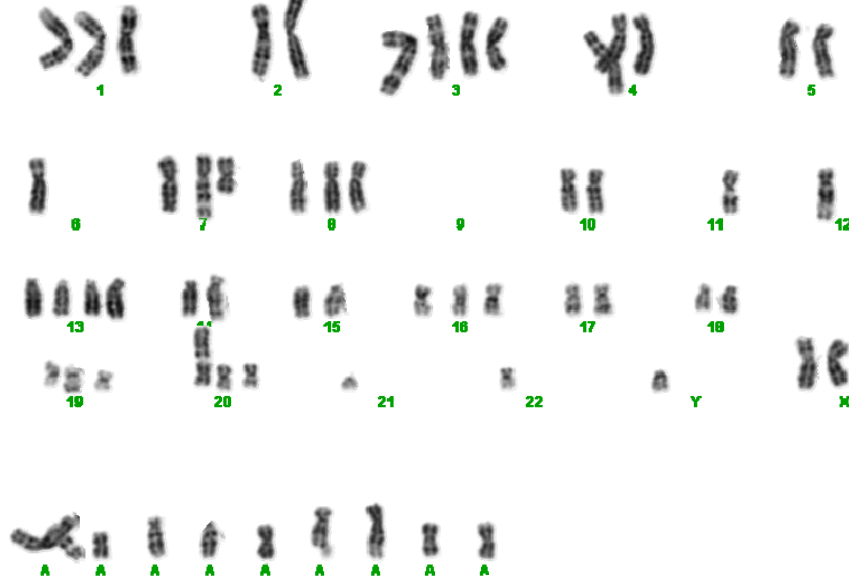
Giving rise to the *BCR1-ABL* fusion gene.

Nowell & Hungerford. Chromosomes of normal and leukemic human leucocytes. J. Natl. Cancer Inst. 1960

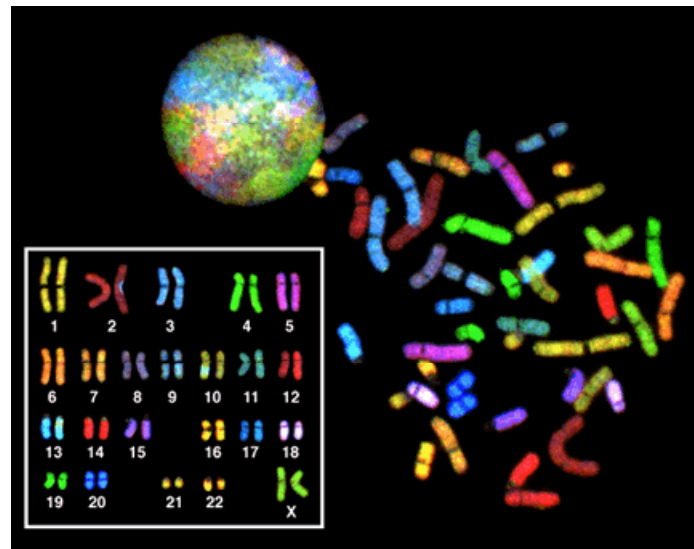
## Complex karyotype (testicular cancer)



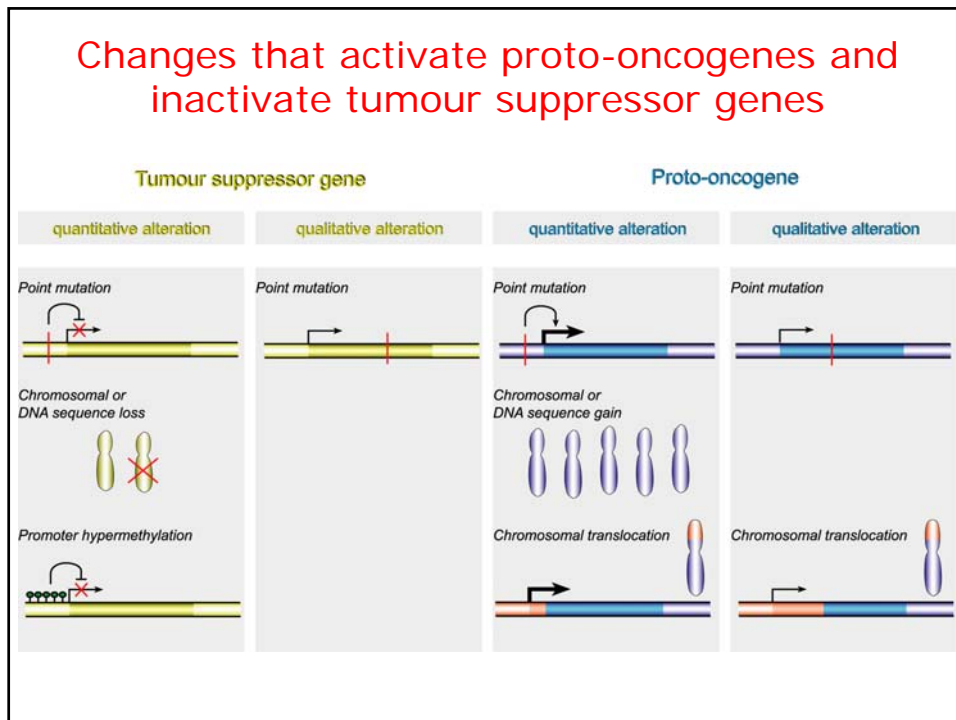
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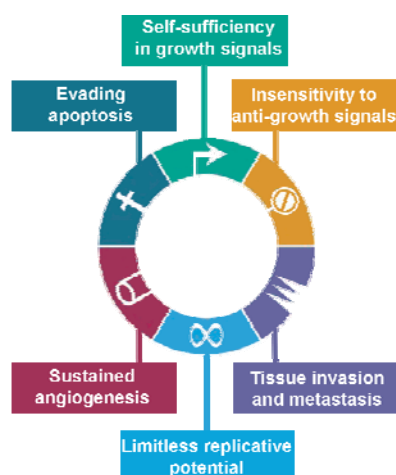
### Spectral karyotype, the metaphase genome



## Changes that activate proto-oncogenes and inactivate tumour suppressor genes

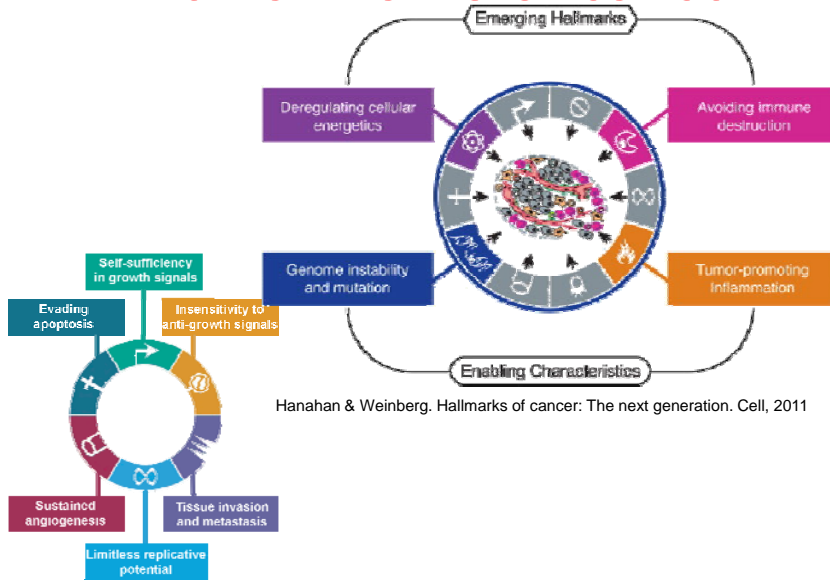


## The hallmarks of cancer

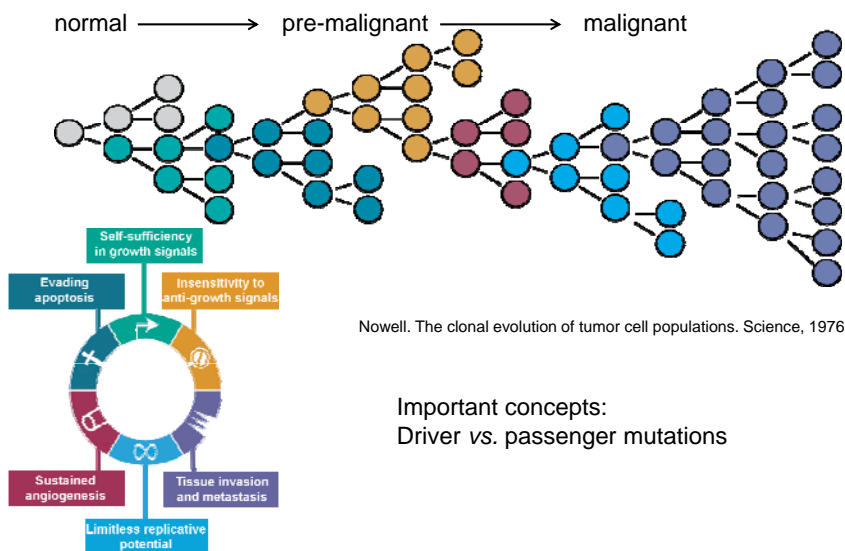


Hanahan & Weinberg. The hallmarks of cancer. Cell, 2000

## The hallmarks of cancer



## Clonal selection



## Tumour suppressor genes

- Protect against malignant phenotype.
- Knudson's two-hit model for inactivation of tumour suppressor genes.
  - Knudson. Mutation and cancer: statistical study of retinoblastoma. Proc. Natl. Acad. Sci. USA, 1971
- Examples of tumour suppressor genes:
  - *RB1*, *TP53*, *CDKN2A*, *PTEN*, *APC*
- Classes of tumour suppressors:
  - Gatekeepers vs. caretakers
  - Mutations in caretaker genes (e.g. DNA repair genes) lead to genomic instability, which again increases likelihood of mutations in gatekeeper genes (and in proto-oncogenes)

## Proto-oncogenes

Conversion from proto-oncogene to oncogene is **dominant**, and oncogenes get hyperactive through **quantitative or qualitative changes**.

- Quantitative: overexpression
  - Gene amplification (*ERBB2*)
  - Chromosomal translocation (*IGH-MYC*)
  - Point mutations with regulatory effects (e.g. in promoter, UTRs)
  - Trans effects: transcription factors, cell signalling, viral integration, etc.
- Qualitative: functional switch
  - Gain-of-function mutation (e.g. point mutation; *KRAS*, *EGFR*)
  - Chromosomal translocation (*BCR-ABL1*)
  - Alternative splicing (*BCL2L1*)

The screenshot shows the Cancer Genome Project website. The main heading is "Cancer Gene Census". Below the heading, there is an "Overview" section with two paragraphs of text. To the right of the text is a table titled "Cancer Gene Census" with two columns: "Sorted By" and "Number".

**Overview**

The Cancer Gene Census is an ongoing effort to catalogue those genes for which mutations have been causally implicated in cancer. The original census and analysis was published in [Nature Reviews Cancer](#) and [supplemental analysis information](#) related to the paper is also available.

The census is not static but rather is updated regularly/as needed. In particular we are grateful to Felix Mitelman and his colleagues in providing information on more genes involved in uncommon translocations in leukaemias and lymphomas. Currently, more than 1% of all human genes are implicated via mutation in cancer. Of these, approximately 90% have somatic mutations in cancer, 20% bear germline mutations that predispose to cancer and 10% show both somatic and germline mutations.

Sorted By	Number
<a href="#">Amplification</a>	11
<a href="#">Chromosome</a>	412
<a href="#">Frameshift mutation</a>	73
<a href="#">Germline mutation</a>	73
<a href="#">Large deletion</a>	29
<a href="#">Missense mutation</a>	105
<a href="#">Nonsense mutation</a>	69
<a href="#">Other mutation</a>	15
<a href="#">Somatic mutation</a>	370
<a href="#">Splicing mutation</a>	46
<a href="#">Symbol</a>	412
<a href="#">Translocation</a>	297

## Strategies for identification of proto-oncogenes

- Transfections of parts of cancer genomes into immortalized mouse-derived fibroblastic cells (gain-of-function; e.g. *RAS*)
- Cytogenetics (e.g. translocations and amplifications)
- RNAi, causing e.g. reduced cell growth or increased apoptosis
- Microarrays/HT-sequencing (combining measurements of DNA copy numbers with gene expression)



## Use of genome-level tools in identification of cancer-critical genes

- Gene expression microarrays
  - gene level
  - exon level
  - fusion transcripts
- DNA copy number microarrays
- Tissue microarrays
- Functional cell microarrays
- High-throughput sequencing (DNA & RNA)

## DNA microarrays, Results and visualisation



GeneNo	sample 1	sample 2	sample 3	sample 4	sample 5	sample 6	sample 7
Gene_1	0.919	0.836	0.795	1.026	0.553	0.788	1.003
Gene_2	2.861	2.483	2.101	1.369	4.683	5.299	0.920
Gene_3	1.582	1.851	1.657	1.144	1.835	1.600	1.043
Gene_4	1.197	1.267	1.287	0.843	0.583	0.870	1.413
Gene_5	0.780	0.695	0.844	1.082	0.540	0.820	3.099
Gene_6	0.745	0.974	0.829	0.735	2.678	0.906	1.073
Gene_7	1.191	0.794	1.175	1.111	0.606	0.688	1.233
Gene_8	1.748	2.741	1.958	0.695	2.079	1.501	0.971
Gene_9	1.118	1.613	0.808	1.311	1.050	1.000	0.756
Gene_10	0.974	1.000	0.900	0.839	0.627	1.196	1.523
Gene_11	0.970	0.865	0.993	1.075	1.202	0.995	1.052
Gene_12	0.731	0.722	0.622	0.817	0.736	0.774	1.000
Gene_13	0.988	1.197	1.144	1.183	0.776	0.718	1.389
Gene_14	1.012	1.072	0.802	1.004	1.325	1.395	1.212
Gene_15	1.625	1.340	2.034	2.008	0.618	0.961	0.873
Gene_16	1.336	2.592	2.164	1.158	0.409	2.510	1.223
Gene_17	4.201	5.179	5.924	0.689	1.624	2.729	0.839
Gene_18	0.946	0.571	0.923	1.522	1.571	1.458	0.371
Gene_19	1.432	1.286	1.556	1.385	0.696	0.890	0.982
Gene_20	1.002	0.790	1.107	1.252	0.594	0.577	0.883

samples →

sample intensity value

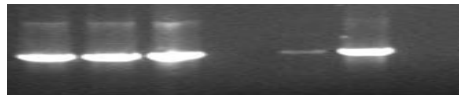
reference intensity value

→ red/green ratio

genes/  
probes ↓



## DNA microarrays, Results and visualisation



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← samples

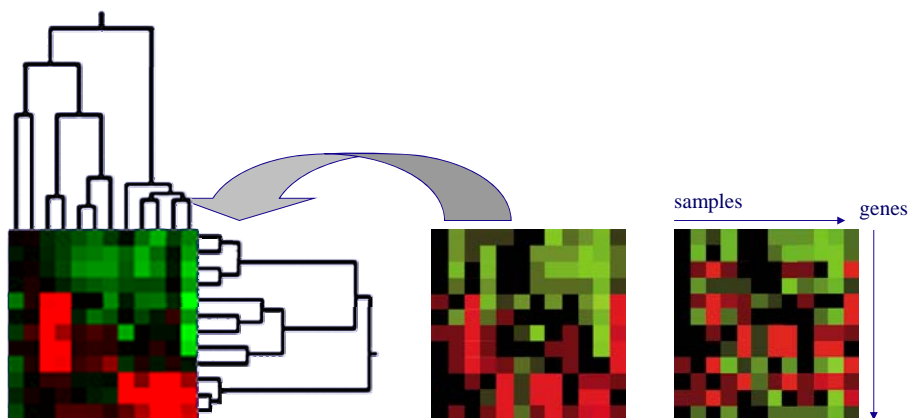
sample intensity value  
reference intensity value

→ red/green ratio

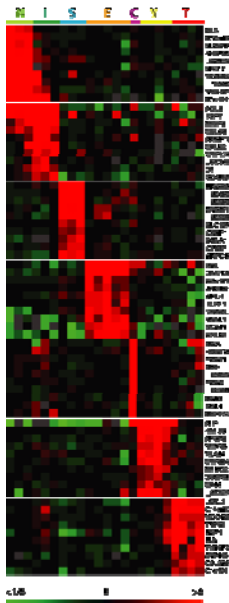
genes/  
probes ↓

## DNA microarrays, Results and visualisation

- Hierarchical cluster analysis

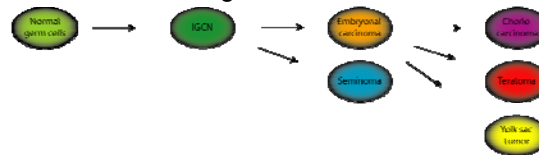


## DNA microarrays, Results and visualisation

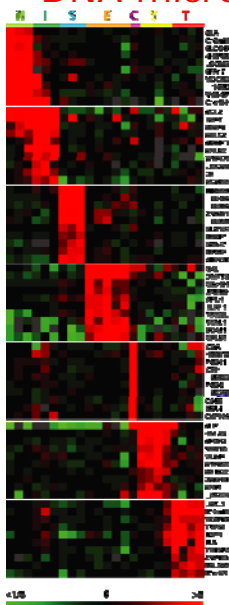


- Hierarchical cluster analysis
- Significance Analysis of Microarrays (SAM)
  - individual analyses of each subtype against the rest of the sample set

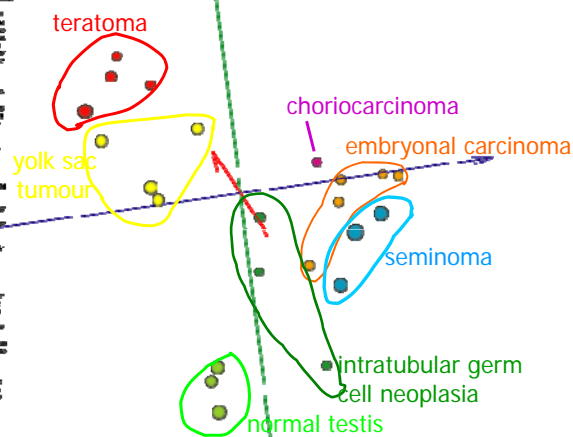
### Testicular tumourigenesis



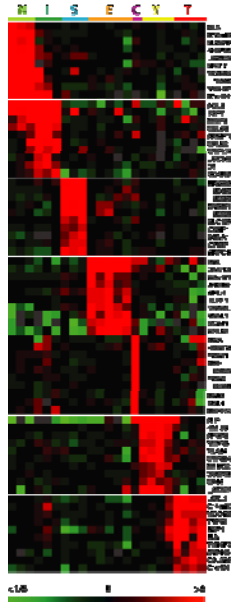
## DNA microarrays, Results and visualisation



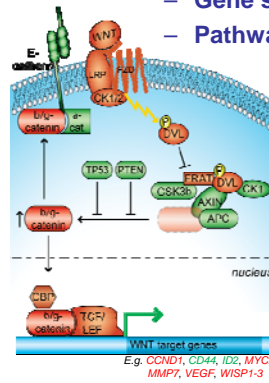
- Hierarchical cluster analysis
- Significance Analysis of Microarrays (SAM)
- Principal components analysis



## DNA microarrays, Results and visualisation



- Hierarchical cluster analysis
- Significance Analysis of Microarrays (SAM)
- Principal components analysis
- Systems biology tools
  - Gene sets enrichments analysis
  - Pathway mapping

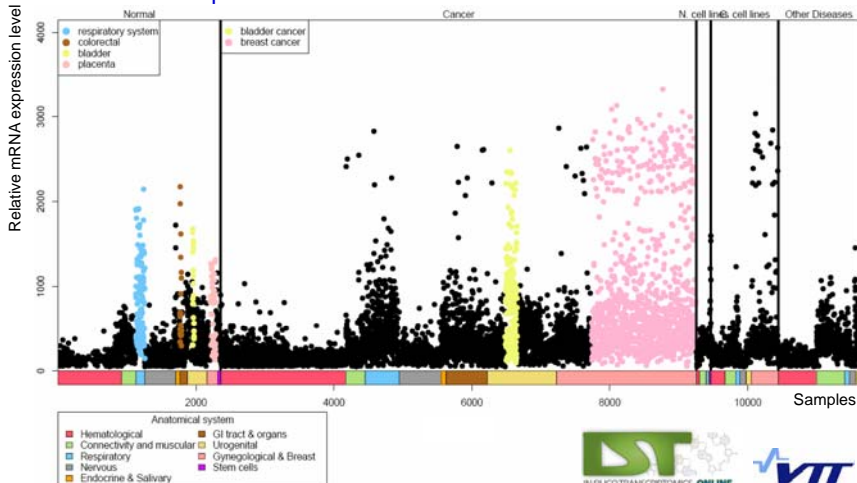


## Gene expression databases

- Data repositories:
  - Gene expression omnibus (GEO) at the National Center for Biotechnology Information (NCBI): [www.ncbi.nlm.nih.gov/geo](http://www.ncbi.nlm.nih.gov/geo)
  - ArrayExpress at the European Bioinformatics Institute (EBI)
- Database interpretation and visualisation
  - Oncomine ([www.oncomine.org](http://www.oncomine.org))
  - MediSapiens ([www.medisapiens.com](http://www.medisapiens.com))

## MediSapiens, visualisation of gene expression across the human sample-space

### ERBB2 overexpression in breast cancer



Herceptin, an antibody against ERBB2 (alias HER2), used to treat breast cancers with amplification of the *ERBB2* gene (17q12)

Kilpinen *et al.*, Genome Biology, 2008

## Transcript variation

<p><b>cassette/exon skipping</b></p>	3316	The most common sources of variation among human transcripts (mostly alternative splicing)  Most expression microarray platforms provide one measurement per gene
<p><b>alternative 5'end (1)</b></p>	2386	
<p><b>retained intron</b></p>	1821	
<p><b>alternative poly-A site (1)</b></p>	1671	
<p><b>alternative 5'end (2)</b></p>	1637	
<p><b>alternative acceptor site</b></p>	1060	
<p><b>alternative donor site</b></p>	788	
<p><b>alternative poly-A site (2)</b></p>	782	

Other: 2429

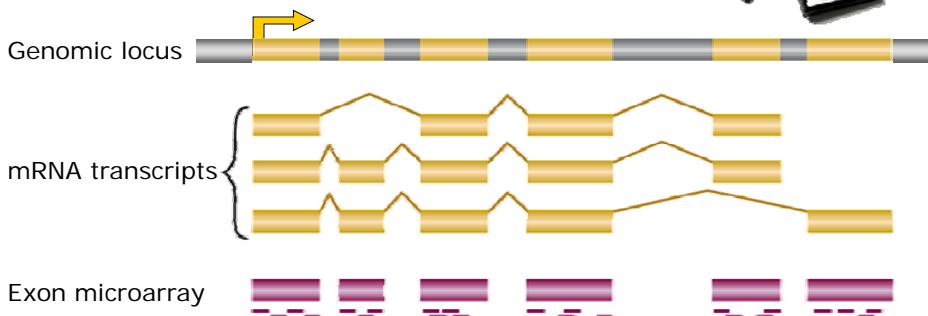
Statistics from ASTRA (Alternative Splicing and TRanscription Archives)  
<http://alterna.cbrc.jp/index.php>

## Affymetrix exon microarrays

Provides gene expression measurements from individual exons

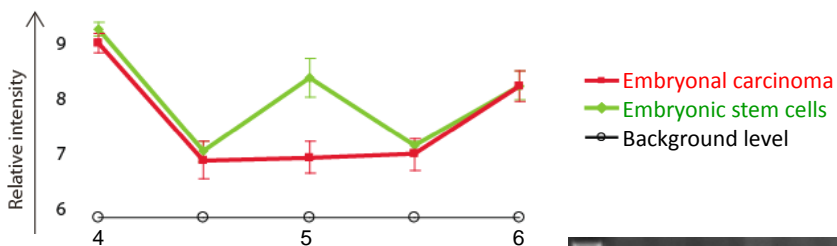
5.4 million probes, measuring

- 325 000 known exons
- ~1 million extra exons based on sequence prediction

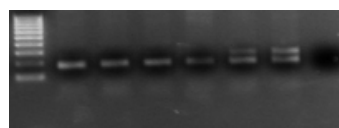
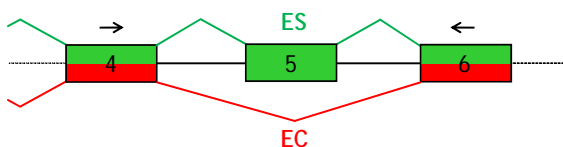


Most DNA microarrays provide one single measurement per gene (in the 3' end)

## Detection of alternative splicing events

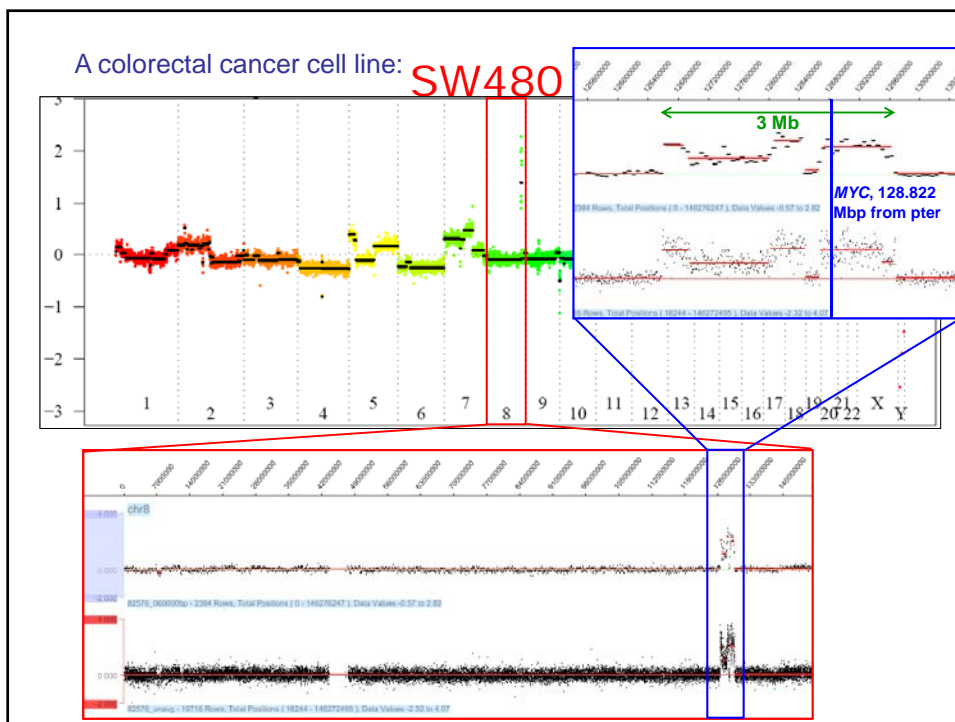
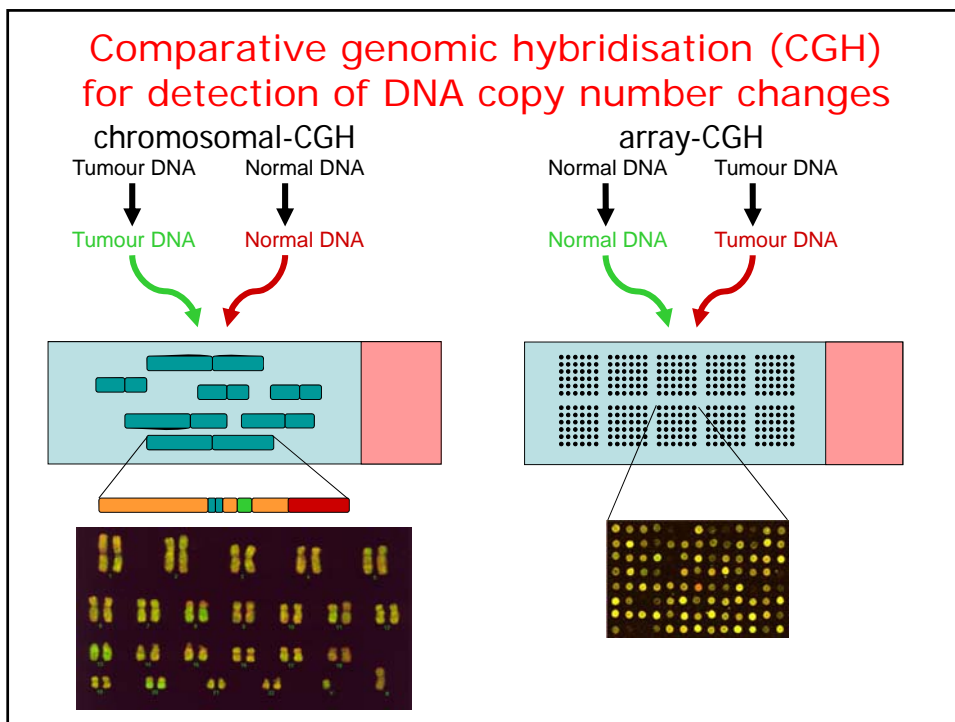


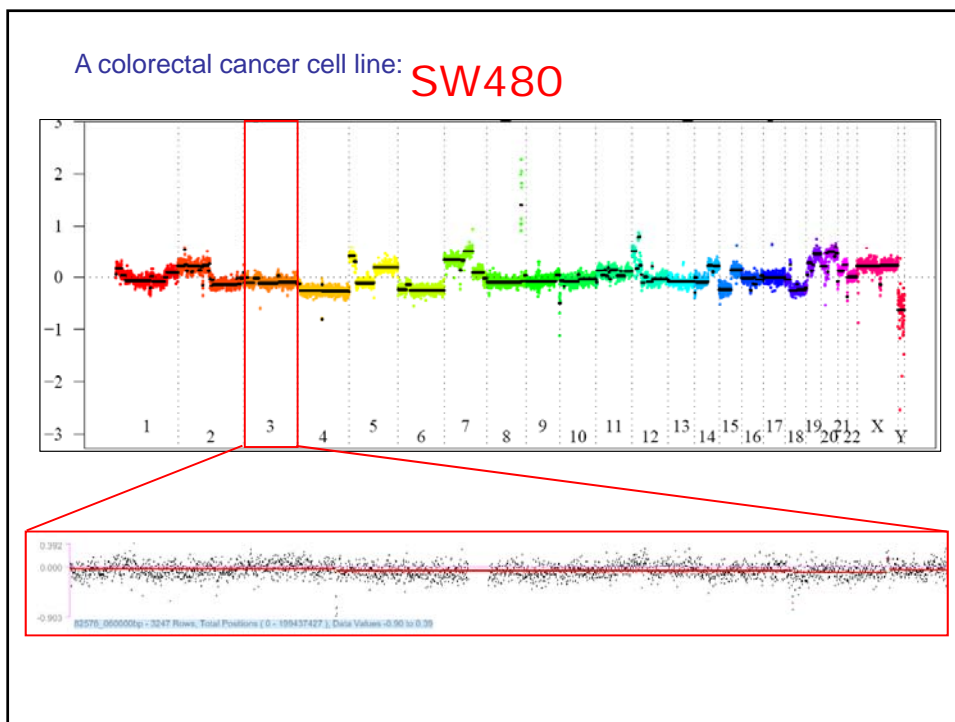
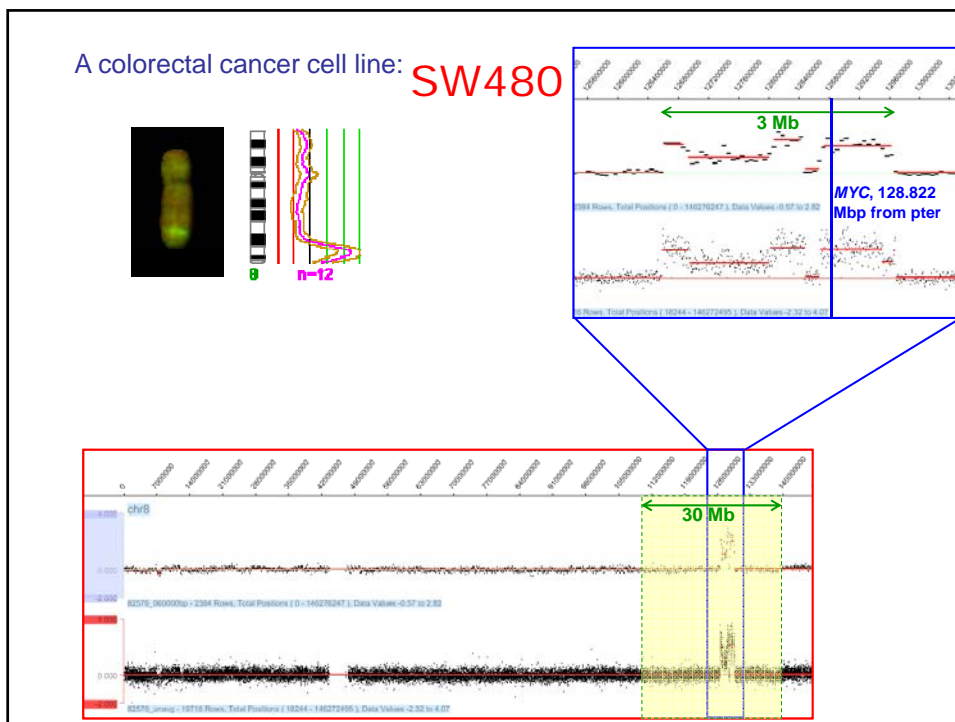
Probes in 5' to 3' direction within a gene encoding a zink finger transcription factor

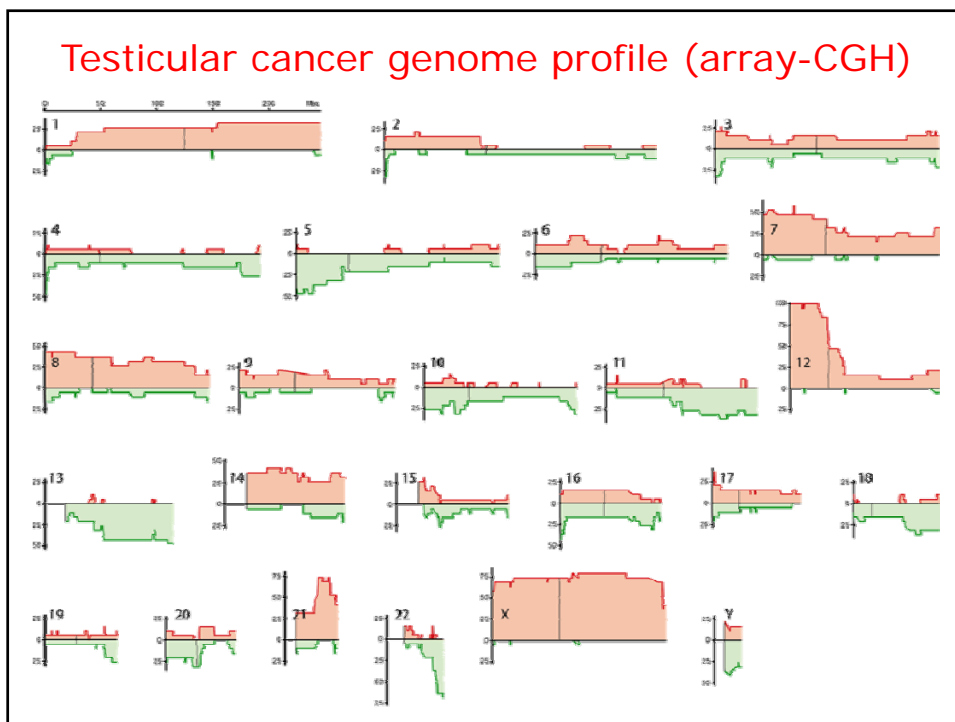
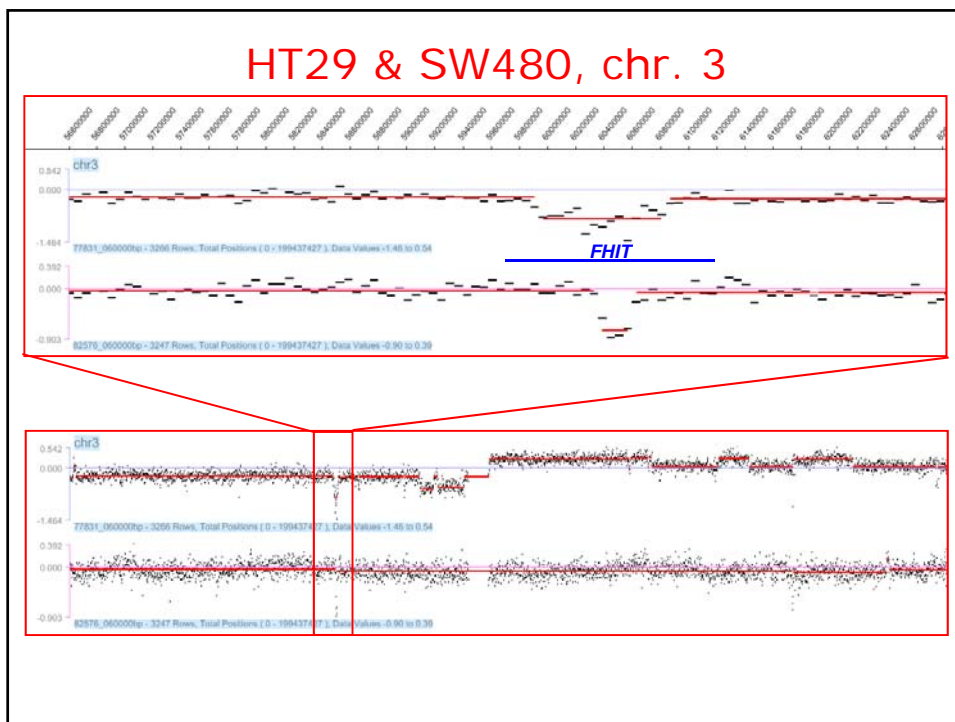


EC ES -

Alagaratnam *et al.*, in prep.

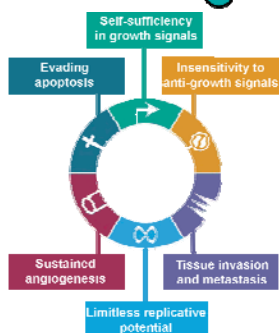
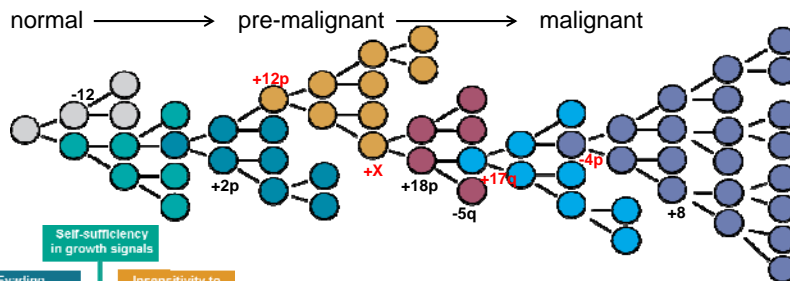








## Clonal selection



Hanahan & Weinberg. The hallmarks of cancer. Cell, 2000

Early events are more easily detected

Important events are more likely to be non-random across a series of cancers. This is because these are events which give rise to selective advantage

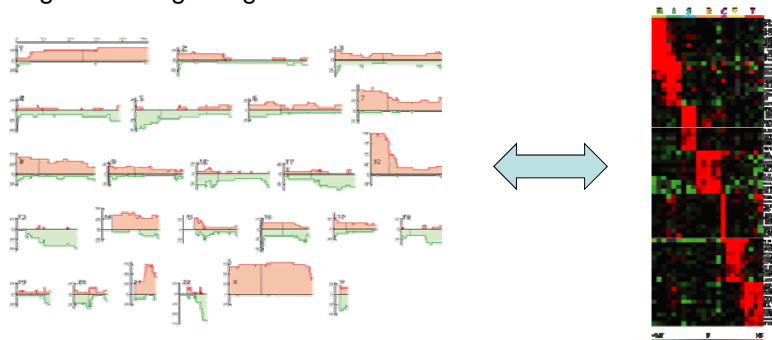
The selective advantage is probably due to one or several genes which expression are altered as a consequence of the DNA copy number change

## Integrated genome and transcriptome analyses

Identification of genes that are over- or under-expressed in samples also having increased or decreased DNA copy number of these genes' loci.

→ potential **proto-oncogenes** or **tumour suppressor genes**

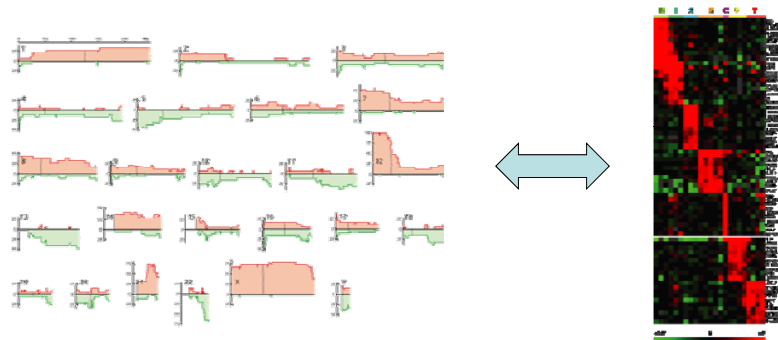
*i.e.*, genes that give the cells selective advantage from having their genomic regions gained or lost.



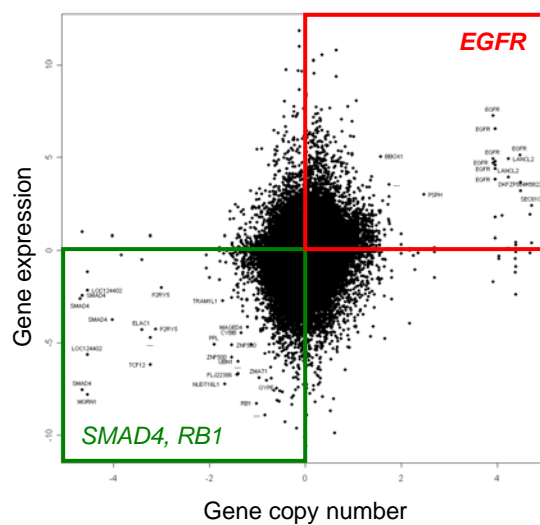
## Integrated genome and transcriptome analyses

For each sample, each gene have two measurements:

- DNA copy number level
  - RNA expression data
- Crude analysis: plot these against each other!



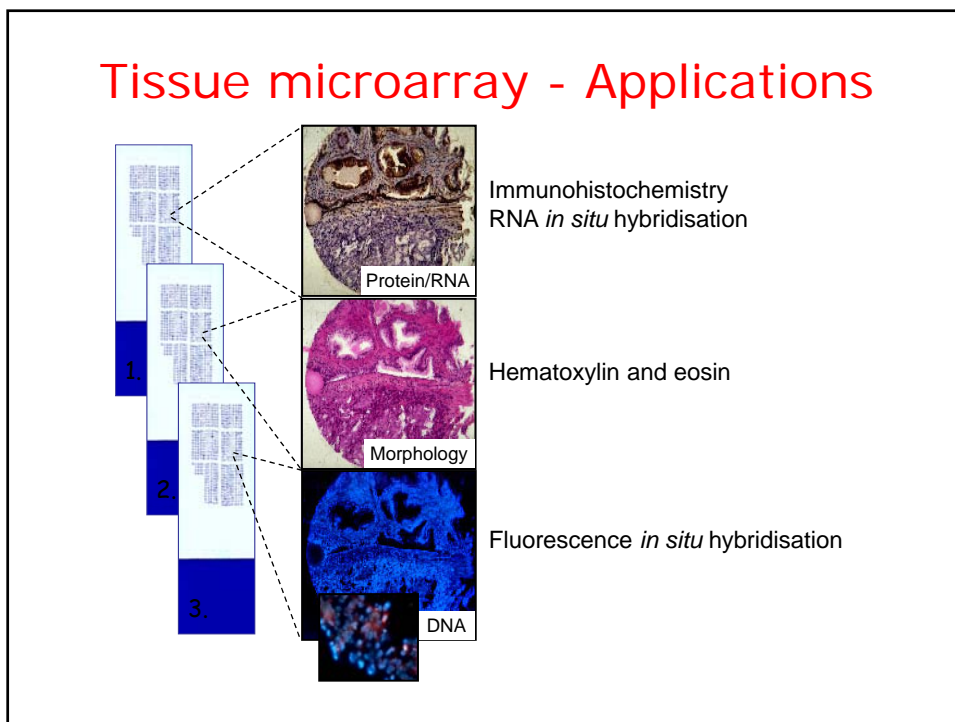
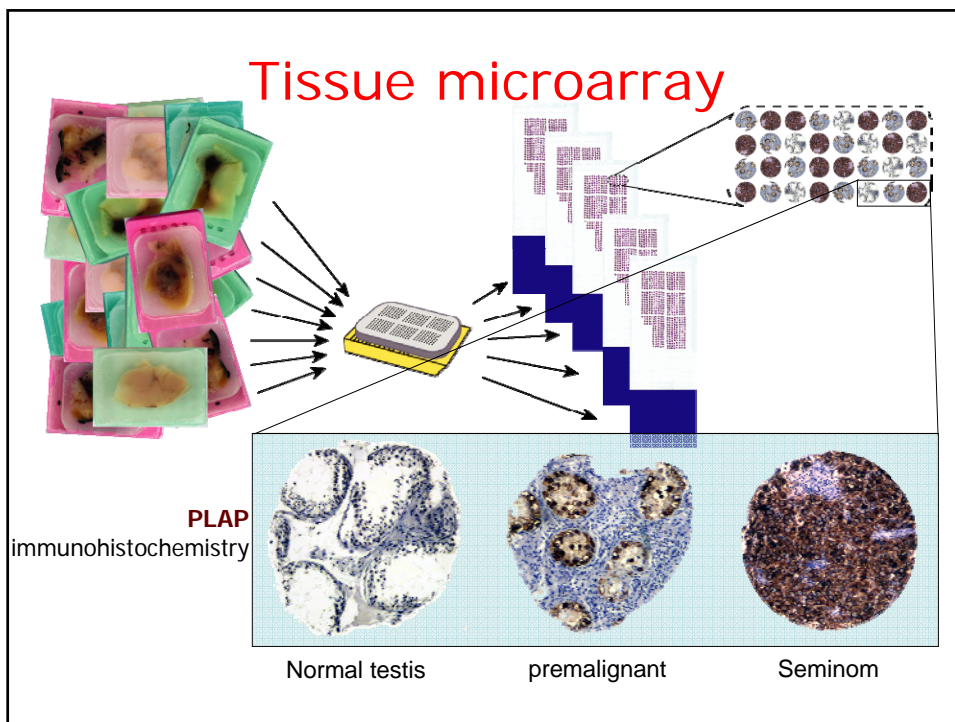
## DNA-RNA outlier analysis



Integration of gene copy numbers with gene expression levels in the same sample

Clear segregation of known key cancer genes, along with a handful of novel genes

Figure from Henrik Edgren *et al.*



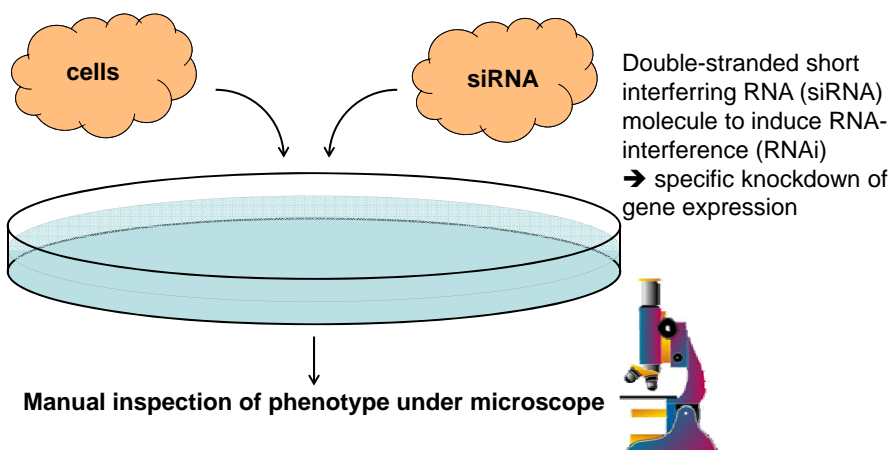
## Tissue microarray - Benefits

- Analyse many tissue samples simultaneously
- Standardisation
  - Minimises experimental errors caused by differing conditions
- “Amplification” of tissue resources
- Saves you time, money, and labour

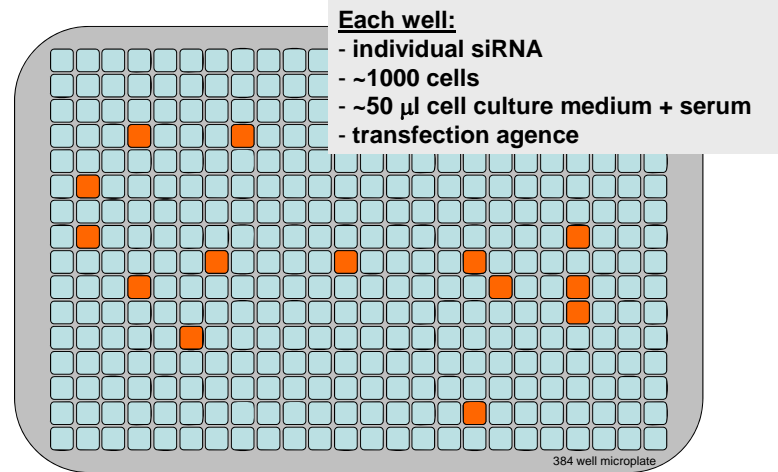
## Applications in cancer research

- Does your candidate gene/protein have prognostic value?
- Can it be used as a diagnostic marker?
- What is the frequency of a molecular alteration in different tumour types?

## siRNA, knockdown of gene expression

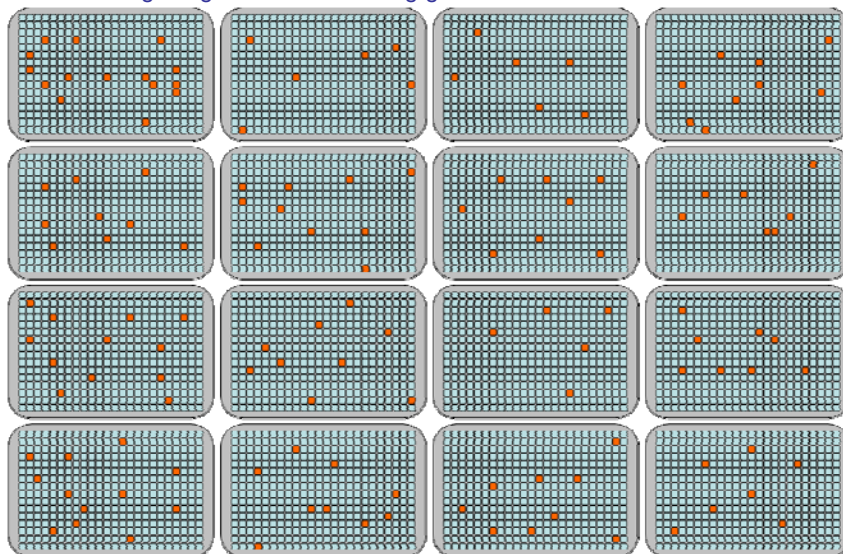


## High-throughput siRNA screening

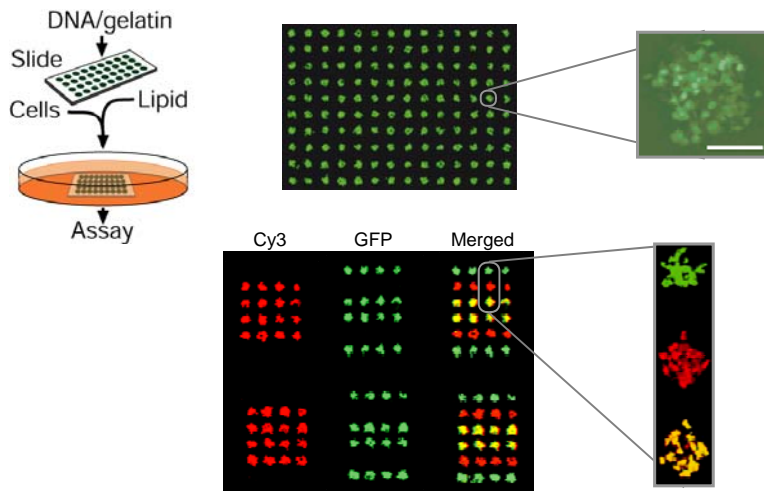


## High-throughput siRNA screening

RNAi goes genomic: elucidating gene function with siRNA libraries



## Functional cell microarray

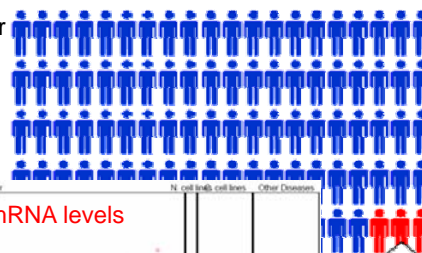


Ziauddin & Sabatini, Nature, 2001

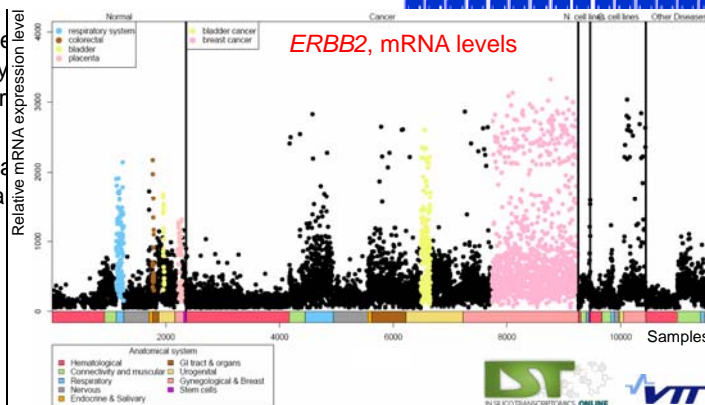
## Cancer Biomarkers

### Lessons from quantitative gene/protein expression

- Genes/proteins highly expressed in cancer are commonly used as biomarkers for diagnostics (e.g. PSA in prostate cancer) and drug sensitivity (e.g. ERBB2, aka HER2, in breast ca.).



- Challenge: Always require...
- Biomarkers...

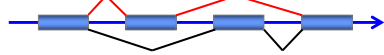


PSA finds these three cases

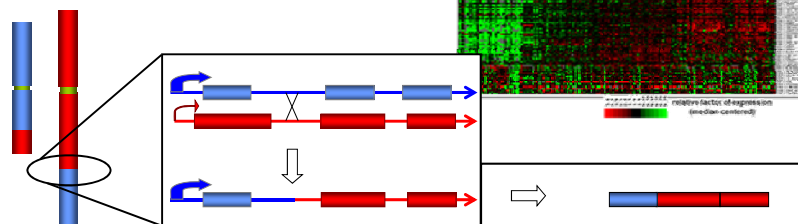


## Strategies to identify novel cancer-specific transcripts

- 1. Embryo-specific transcripts
- 2. Specific transcript processing variants

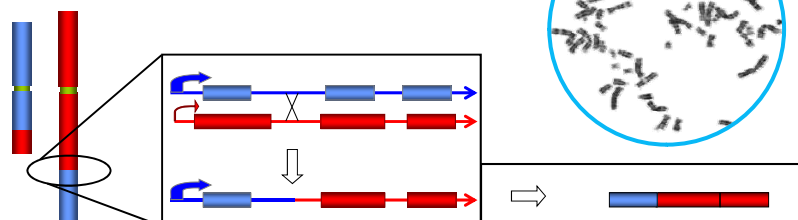


- 3. Fusion genes



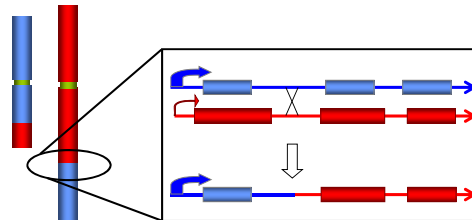
## Fusion genes in cancer

- Caused by e.g. chromosomal translocations, deletions, and inversions
- Particularly common in haematological cancers, sarcomas, and also in certain carcinomas (e.g. prostate, lung, thyroid)
- Identification of certain fusion genes are currently performed for differential diagnosis or therapeutic decision-making
- Several technological limitations

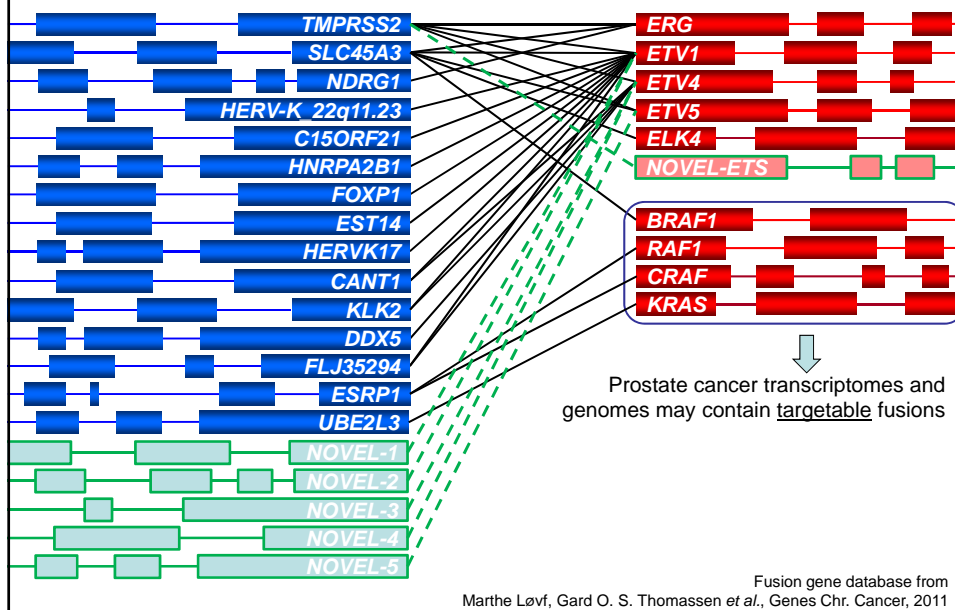


## Translocations and fusion genes

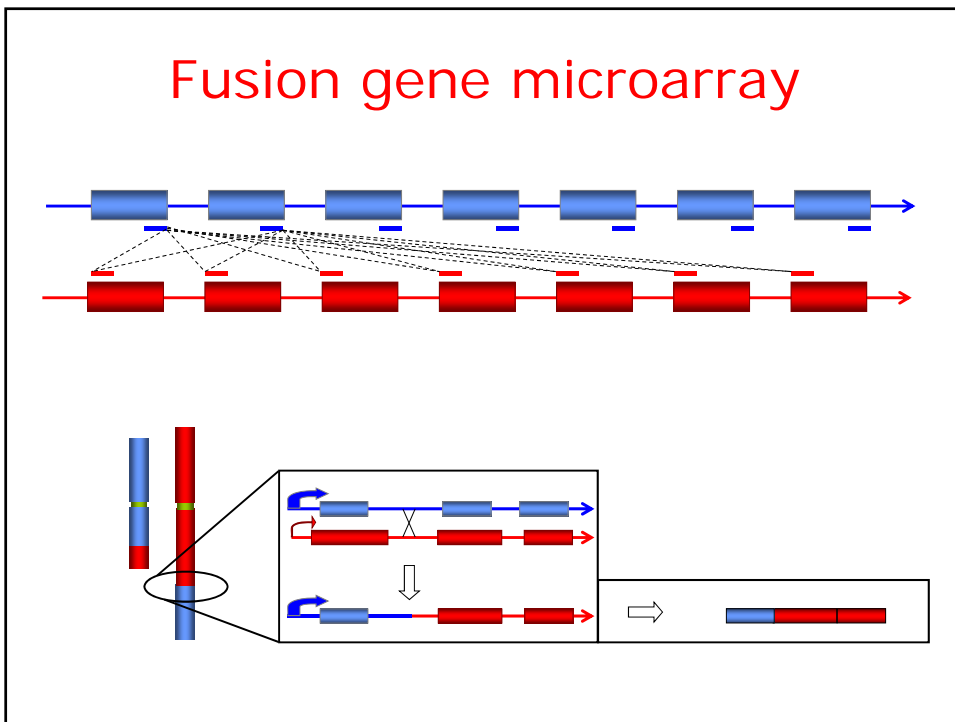
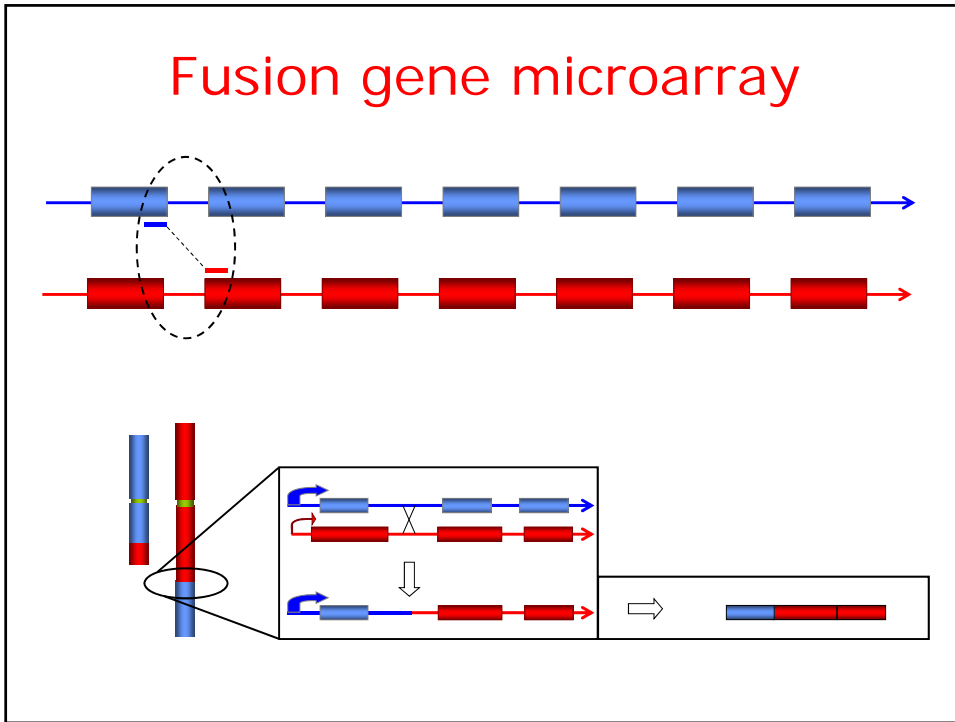
- Chronic myelogenous leukaemia, t(9;22)(q34;q11)
  - Philadelphia chromosome, first identified human translocation
  - *BCR-ABL1*, encoding a fusion protein with domains from both original genes, including a tyrosine kinase (TK) activity.
  - Gleevec binds at the kinase domain, and inhibits phosphorylation of TK target proteins.
- Burkitt's lymphoma, t(8;14)(q24;q32)
  - *MYC* proto-oncogene juxtaposed with the immunoglobulin heavy chain gene: *IGH-MYC*



## Fusion genes in prostate cancer

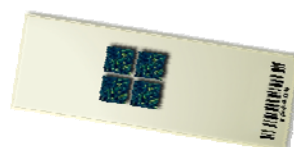




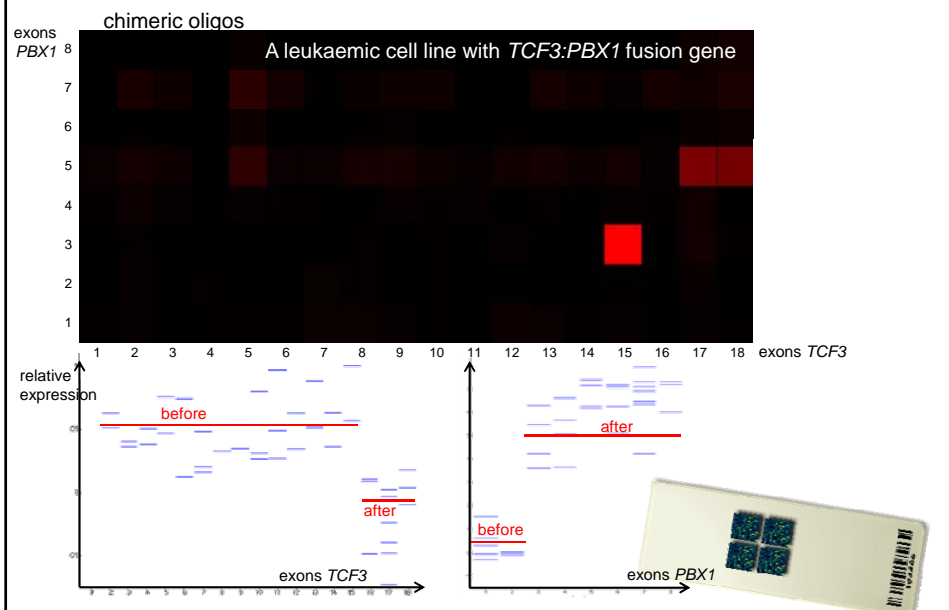


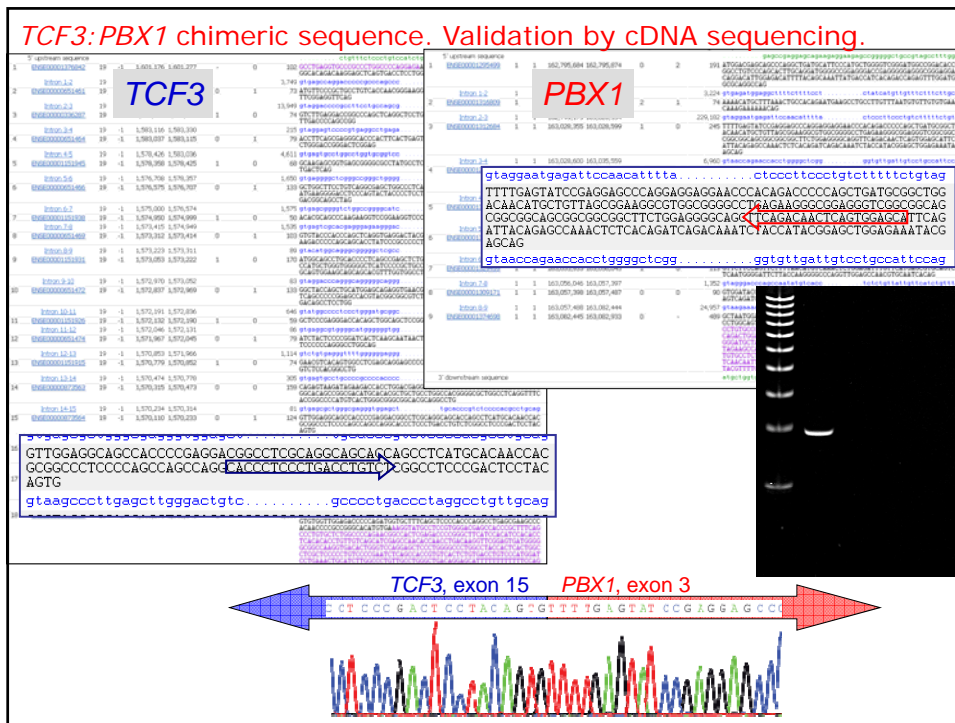
## Pilot design, fusion gene microarray

- Databases/literature
  - 275 known fusion genes at time of pilot array design
- Sequences and exon annotation from Biomart.org
- Generation of chimeric sequences (~60 000)
  - Automised by script programmed in Python
- Oligo design
  - 34-40mers, variable length for isothermic properties
  - Chimeric oligos with matching Tm from up- and downstream fusion partners
  - Intragenic oligos
- Microarray platform:



## Visualisation of results





Pilot: <http://www.molecular-cancer.com/content/8/1/5>

## Fusion gene microarray, version 2.1

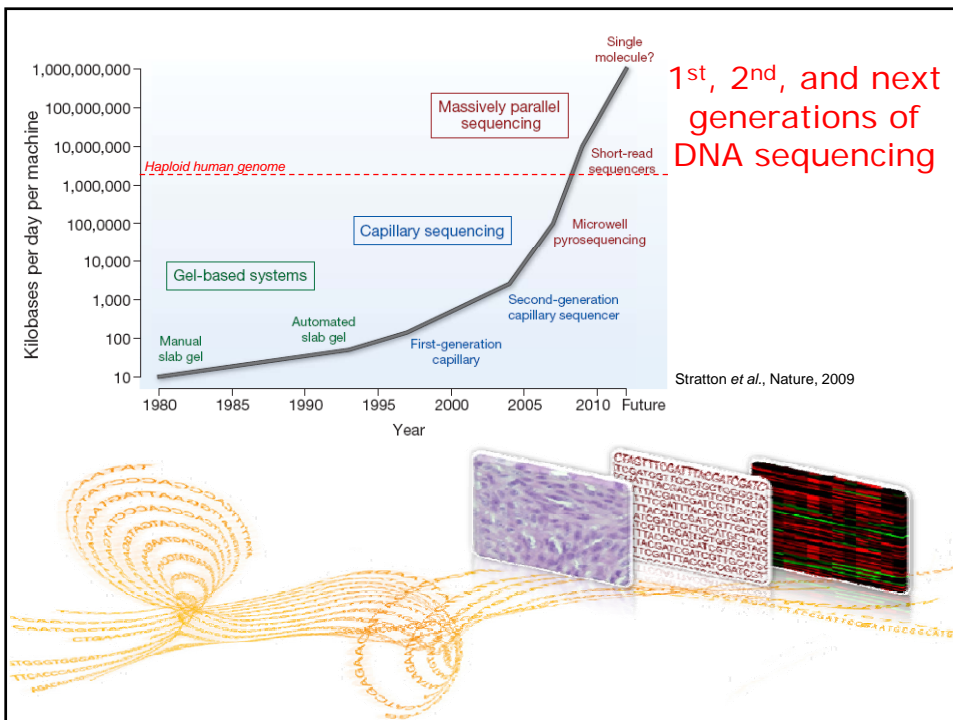
- 556 gene pairs, including candidate fusion genes from high-throughput sequencing
- NimbleGen 3 x 720k microarray format
- Discovery/validation
  - Known and putative fusion genes in solid tumours
- Diagnostic potential
  - leukaemia, sarcomas, other solid tumours?

Leff, Thomassen *et al.*, Genes Chromosomes and Cancer, 2011

## Next generation sequencing

New sequencing methodologies that do not use the Sanger sequencing methodology (di-deoxy sequencing), and which have unprecedented high throughputs (giga-base level)

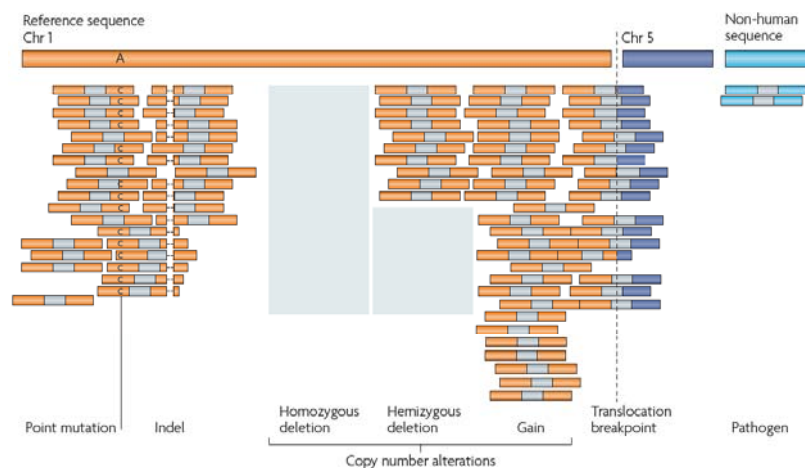
*454, SOLiD, GA-IIx, HiSeq-2000, Helicos, PacBio, Ion Torrent, Dover (Polonator), & Complete Genomics*



## Main applications

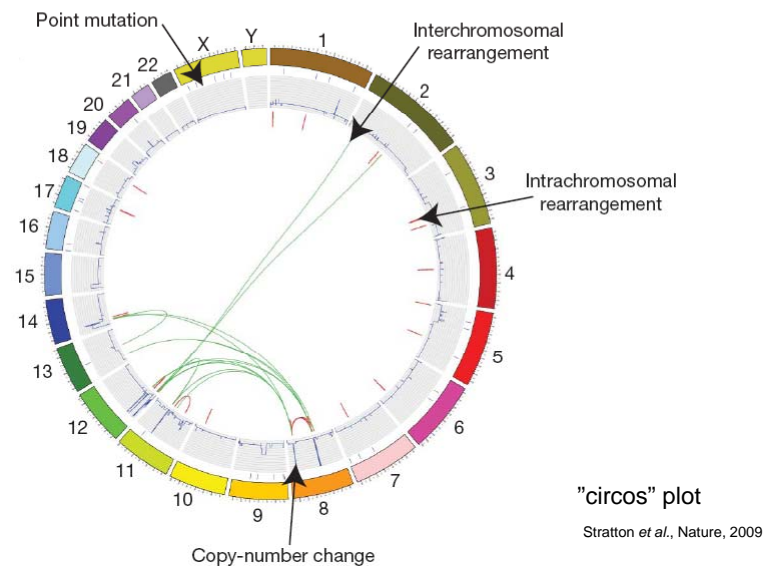
- **Genome** analysis, DNA-seq
  - Variant detection (mutation/polymorphism)
    - base level
    - structural changes (paired end)
  - Copy number analysis
- **Transcriptome** analysis, RNA-seq
  - Expression levels (tag seq)
  - Transcript variants (paired end)
  - Small RNA analysis (e.g. miRNA profiling)
- **DNA-protein** interactions
  - ChIP-seq (chromatin immunoprecipitation)
  - Transcription factor binding sites

## Types of genome alterations detectable by next-generation sequencing



Meyerson *et al.*, Nat.Rev.Genet., 2010

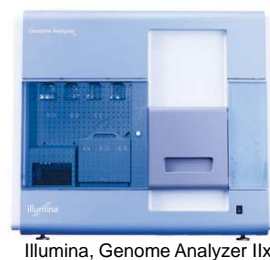
## Visualising a cancer genome



## Sequencing based transcriptome analysis (RNA-seq)

Characterize all transcriptional activity, coding and non-coding, in cell population without *á priori* assumptions

- Novel transcript structures from aberrant splicing, promoter usage, and fusion genes
- digital expression levels/relative abundance of transcripts
- mutation detection
- non-coding/regulatory RNAs



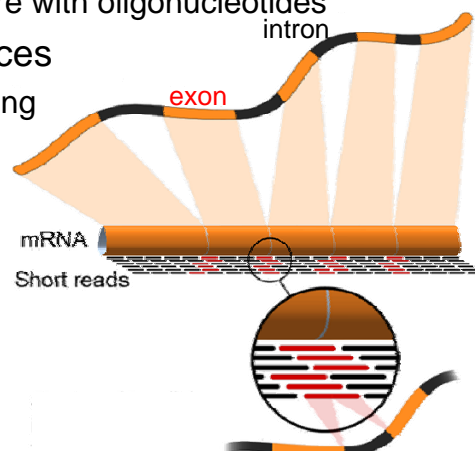
## Unbiased

- CHIP-seq, not limited to known promoter areas
- RNA-seq, not limited to known transcript structures and exons
- DNA-seq, mutation analysis not limited to annotated exons

## When biased is the goal

*E.g.* detection of mutations within

- a specific gene or genomic locus
  - long range PCR, capture with oligonucleotides
- all transcribed sequences
  - transcriptome sequencing
  - exome-capture









## Acknowledgements

The presented research data have been obtained in projects carried out at **Dept. Cancer Prevention**, Inst. Cancer Res., Radium Hosp., Oslo University Hosp.



[www.rr-research.no/cancerprevention](http://www.rr-research.no/cancerprevention)